



ORIGINAL ARTICLE

Amino Acids, Solubility, Bulk Density and Water Holding Capacity of Novel Freeze-Dried Cow's Skimmed Milk Fermented with Potential Probiotic *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6



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Received 8 May 2021; accepted 23 June 2021

Available online 25 June 2021

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Peer review under responsibility of King Saud University.



KEYWORDS

Lactobacillus;
Freeze-drying;
Skim milk;
Amino acids;
Solubility index;
Bulk density

Abstract *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6 showed a potential probiotic role *in vivo* and *in vitro* in a preliminary unpublished work. This study demonstrated that *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6 mediated modifications of the physicochemical properties of fermented skimmed cow's milk. Skim milk was inoculated with 1% (w/v) of *Lactobacillus plantarum* Bu-Eg5 (T₁), 1% (w/v) of *Lactobacillus rhamnosus* Bu-Eg6 (T₂) and 1% (w/v) of 1:1 *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6 (T₃). After that, fresh skimmed milk control (C), T₁, T₂, and T₃ were freeze-dried (Zirbus Technology model: VaCo 5-D, S/N: COM98754, Germany). The resulting freeze-dried fresh and fermented milks powders were evaluated for their solubility index, bulk density and water holding capacity (WHC). The amino acids compositions of the freeze-dried treatments were analyzed using High Performance Amino Acid Analyzer (Biochrom 30). The microbiological quality of freeze-dried fresh and fermented milks powders was measured by enumerating lactic acid bacteria, yeast & mould and coliform bacteria. The total protein content of fresh freeze-dried skimmed milk (38.69%) was higher than that of fermented freeze-dried skimmed milk powders. The bulk density was 16.67 g/mL for all treatments. The solubility of fresh freeze-dried skimmed milk was the highest (100%), followed by the other treatments (78.95%). The WHC of fresh freeze-dried skimmed milks (11.16%) was lower than the other samples. The highest total essential amino acids were observed in fresh freeze-dried skimmed milk sample (17.26%) followed by the freeze-dried skimmed milk fermented with *Lactobacillus plantarum* Bu-Eg5 (16.22%) while the lowest recorded in the freeze-dried skimmed milk sample fermented with 1:1 *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6 (12.87%), there was a very low bacterial count in the control treatment (1.09 log CFU/ml). Yeast & mould and coliforms were not detected in all samples.

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1. Introduction

Understanding of the good diet today is not only to supply the body with the right quantities of nutrient components, but also, to play a physiological role against different diseases (Domínguez Díaz et al., 2020).

Fermented milks are broadly manufactured in numerous nations. This kind of procedure is probably the most seasoned technique utilize to broaden the shelf life of usability of milk and drilled by individuals for millennia (Pederson, 1971).

Fermented milks have a chemical composition which gives a helpful sign of the expected health benefit of these items. The primary parts are protein, fat, sugar, minerals, and nutrients. Be that as it may, the bioactive peptides ingredients (eg. immuno-peptides, casomorphins, α - and β -lactorphan, phosphor peptides or lactoferricin), which are professed to be well-being improving segments, ought not be disregarded (Meisel & Bockelmann, 1999).

The helpful wellbeing characteristics of fermented milks and their utilization in the Therapy of body infirmities traces all the way back to not many thousands years prior; they have been likewise referenced in Biblical sacred writings, and some old researchers have endorsed it as medication for relieving metabolic issues of the digestive tract and stomach (Oberman & Libudzisz, 1998).

Probiotics were defined as “live microorganisms which when administered in adequate amounts confer a health benefit on the host” (FAO/WHO, 2002)

Probiotics were added to many kinds of foods, mainly cheese, yoghurt and other fermented milks. In the past few decades, the consumption of foods containing probiotics

increased gradually to become a billion dollars industry worldwide (Suez et al., 2019).

Several isolates of lactic acid bacteria were classified as probiotics. The selection probiotics criteria should be differentiated as the ability to live through its way in the digestive system, and at reduced pH, bile salts and enzymes, and the rate of surviving must be not less than 106 CFU/g (Hosseini et al., 2009).

The freeze-drying technique is commonly successfully used to provide ready to preserve of cultures and to use highly concentrated starter cultures (Martos et al., 2007; Velly et al., 2014; Gul et al., 2020)

This technique entails freezing and drying the aqueous solution containing cells in order to remove water by ice sublimation, likely ice crystal formation, and DNA denaturation due to osmolality, resulting in damaged cells (Conrad et al., 2000; Han et al., 2018).

Several studies have revealed that the utilization of skimmed milk to decrease cell damage due to the low temperatures during freeze-drying treatment and after freezing. The Freeze-drying technique of skim milk has an important positive effect due to reducing cellular cell viability by protecting cells during freeze-drying via a viscous layer which resulted by milk proteins on the surface of cells and stabilizing cell membrane. Hence, hydroxyl or amino groups that can replace water on the proteins molecules and membrane lipids (Hubalek, 2003; Carvalho et al., 2004; Chen et al., 2015; Lu et al., 2017; Stefanello et al., 2019; Niu et al., 2016).

The aim of this project was to understand the effects of adding *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6 on gross chemical composition, solubility

index, bulk density, water holding capacity (WHC), and amino acids of fermented freeze-dried cow's skimmed milk.

2. Materials and methods

2.1. Materials

2.1.1. Raw milk

Fresh cow's milk was obtained from the herd of the Faculty of Agriculture, Moshtohor, Benha University, Egypt. The chemical composition of milk was 3.4% fat, 3.2% protein, 5% lactose and 0.61% ash.

2.1.2. Lactic acid bacterial strains

Lactobacillus plantarum Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6 were obtained from the culture collection of Dairy Science Department, Faculty of Agriculture, Moshtohor, Benha University, Egypt. These strains were activated three times on MRS broth medium (HIMEDIA, Pvt. Ltd, India), then transferred to heat treated skim milk (121 °C/5 min) at a rate of 1% and incubated at 37 °C until coagulation.

2.1.3. Preparation of fermented milks

Cream was separated from cow's milk in the dairy pilot plant of the Faculty of Agriculture, Moshtohor, Benha University and yielded cream with 22% fat and skim milk with 0.05% fat. The skimmed milk was divided into 4 portions and filled into sterile glass containers (1L each) then heat treated at 121 °C for 5 min. in the autoclave. Heat treated skim milk was allowed to cool to room temperature, then 4 portions were marked as follows: The 1st portion is control (C), the 2nd portion was inoculated with 1% (w/v) of *Lactobacillus plantarum* Bu-Eg5 (T₁), the 3rd portion was inoculated with 1% (w/v) of *Lactobacillus rhamnosus* Bu-Eg6 (T₂) and the 4th portion was inoculated with 1% (w/v) of 1:1 *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6 (T₃). The inoculation was carried out under aseptic conditions. The inoculated skim milk was incubated at 37 °C until coagulation (~24 h). The resulting fermented milks were evaluated for their microbiological properties by counting lactic acid bacteria on MRS (HIMEDIA, Pvt. Ltd, India), yeast & moulds were counted on potato dextrose agar and coliforms were counted on violet red bile agar media.

2.2. Methods

2.2.1. Freeze-drying of skimmed milk

Freeze-drying of fresh and fermented skimmed milk was done using freeze dryer (Zirbus Technology, S/N: COM98754, model: VaCo 5-D, Germany) at -40 °C under vacuum at a minimum pressure of 0.011 kPa Fig. 1. The operation of freeze drying was completed in 72 h according to the methods described by Ismail et al., (2020) and the freeze-dried skimmed milk was weighed before and after freeze drying to calculate the yield of freeze-dried fermented milk Fig. 2 according to the following Eq. (1):

$$\text{Yield}(\%) = \frac{\text{Weight of freeze dried fermented milk}}{\text{Weight of traditional fermented milk}} \times 100 \quad (1)$$

2.2.2. Solubility index of freeze-dried of fresh and fermented skimmed milk powder

Powder solubility was examined according to the method mentioned by as following Eq. (2) according to the method mentioned by Ismail et al., (2020)

$$\text{Solubility} (\%) = \frac{\text{Supernatant volume}}{\text{Total volume}} \times 100 \quad (2)$$

2.2.3. Bulk density of freeze-dried of fresh and fermented skimmed milk powder

Bulk density of freeze-dried of fresh and fermented skimmed milk powder was examined according to the method mentioned by Akash and Yugal (2017) and calculated as following Eq. (3)

$$\text{Bulk density} = \frac{\text{Powder sample in gram}}{\text{Rise in volume}} \quad (3)$$

2.2.4. Water holding capacity (WHC)

The water holding capacity (WHC) was determined according to the method of Akahn et al. (2012) as follows: 20 g of freeze-dried of fresh and fermented skimmed milk powder were centrifuged at 5000g and 20 °C for 10 min.

The water holding capacity was calculated as following Eq. (4):

$$\text{WHC} (\%) = \frac{\text{Freeze dried fermented milk weight} - \text{Separated Whey weight}}{\text{Freeze dried fermented milk weight}} \times 100 \quad (4)$$

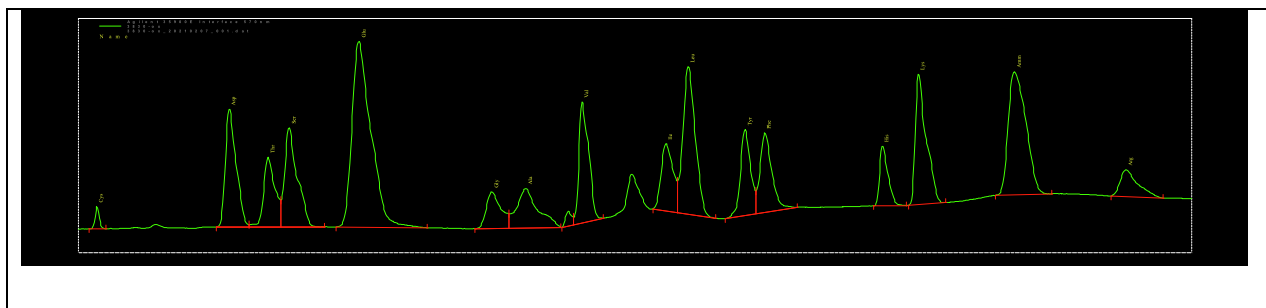


Fig. 1 Chromatogram of amino acids content of freeze-dried fermented milks with *Lactobacillus*.

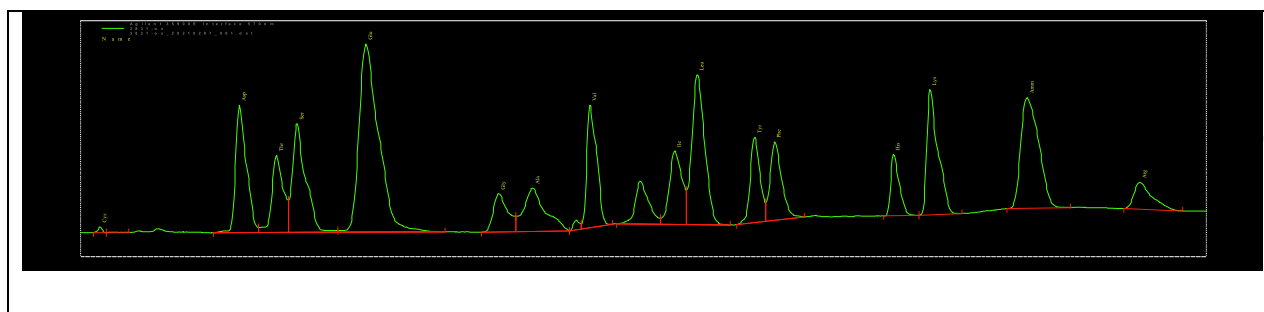


Fig. 2 Chromatogram of amino acids content of freeze-dried fermented milks with *Lactobacillus plantarum* Bu-Eg5.

2.2.5. Gross chemical composition of freeze-dried skimmed milk treatments

Gross chemical composition (moisture, protein, fat and ash) of freeze drying skimmed milks were determined according to AOAC (2019). The carbohydrate content was calculated by difference.

$$\% \text{ Carbohydrates} = 100 - (\% \text{ Protein} + \% \text{ Moisture} + \% \text{ Ash} + \% \text{ Lipids})$$

2.2.6. Determination of amino acid composition

Amino acids composition of freeze-dried fresh and fermented skim milk powders analyzed by (Biochrom 30), hydrolysis were estimated in conical flask via ten milligram of protein was added and 5 ml. of per formic acid was added and the mixture placed in bath for 16 h then sodium disulfite to the oxidized the mixture, the flask adjust the pH to 2.20 by hydroxide solution, the samples injected in Biochrom 30 column according to the method described by Aly et al., (2020) at Food Tech. Res. Inst., Agric. Res. Center, Giza, Egypt. The uncertainty represents an expanded uncertainty (Typ A) expressed at approximately the 95% confidence level using a coverage factor of $k = 2$.

2.2.7. Microbiological analysis

Lactic acid bacteria were analyzed according to standard procedures as mentioned by Ismail et al., (2020a) Yeast and mould counts and coliform counts were computed according to standard procedures Ismail et al., (2020b) at the Faculty of Agriculture, Moshtohor, Benha University, Egypt.

2.2.8. Statistical analysis

Statistical analysis of data was analyzed statistically as mean \pm SD of triplicate according to Aly et al., (2021).

3. Results and discussion

3.1. Gross chemical composition of freeze-dried skimmed milk treatments

Data revealed in Table 1 shows the gross chemical composition of freeze-dried fresh and fermented skim milks. The total protein contents were higher in fresh freeze-dried skimmed milk than in fermented freeze-dried skimmed milks. These values reached 38.69% for control sample (fresh freeze-dried skimmed milk) and recorded 36.75%, 34.42% and 32.47% for skimmed milk fermented with *Lactobacillus plantarum* Bu-Eg5 (T₁), freeze-dried skimmed milk fermented with *Lactobacillus rhamnosus* Bu-Eg6 (T₂) and freeze-dried skimmed milk fermented with 1:1 *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6 (T₃). This decrease in protein content of fermented freeze-dried skimmed milk might be attributed to its use during *Lactobacillus* bacteria growth. Total fat, moisture, total carbohydrates, and ash contents did not changed in all examined treated variants (see Table 2).

3.2. Yield of fresh freeze-dried skimmed milk treatments

The evaluation of freeze-dried skimmed milks powder produced after the freeze-drying process is shown in Table 3.

Table 1 Gross chemical composition of freeze-dried skimmed milks fermented with *Lactobacillus*.

Gross chemical composition	Treatments			
	C	T ₁	T ₂	T ₃
Moisture	5.33 \pm 0.76	5.30 \pm 0.10	5.40 \pm 0.40	5.20 \pm 0.10
Protein	38.69 \pm 0.89	36.75 \pm 0.58	34.42 \pm 0.58	32.47 \pm 0.34
Fat	1.32 \pm 0.13	1.33 \pm 0.15	1.27 \pm 0.25	1.30 \pm 0.10
Ash	8.10 \pm 0.17	8.44 \pm 1.10	8.81 \pm 0.58	8.84 \pm 0.50
Carbohydrates	46.55 \pm 0.19	48.18 \pm 0.12	50.11 \pm 0.13	52.19 \pm 0.15

Values are means \pm standard deviation. C: Control (Fresh freeze-dried skimmed milk); T₁: skimmed milk fermented with *Lactobacillus plantarum* Bu-Eg5; T₂: freeze-dried skimmed milk fermented with *Lactobacillus rhamnosus* Bu-Eg6; T₃: freeze-dried skimmed milk fermented with 1:1 *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6.

Table 2 Yield of freeze-dried skimmed milks fermented with *Lactobacillus*.

Treatments	Yield (%)
C	14.88 ± 0.47
T ₁	15.25 ± 0.59
T ₂	14.68 ± 0.26
T ₃	14.99 ± 0.66

Values are means ± standard deviation. C: Control (Fresh freeze-dried skimmed milk); T₁: skimmed milk fermented with *Lactobacillus plantarum* Bu-Eg5; T₂: freeze-dried skimmed milk fermented with *Lactobacillus rhamnosus* Bu-Eg6; T₃: freeze-dried skimmed milk fermented with 1:1 *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6.

The yield of freeze-dried skimmed milks powder produced ranged from 14.68% in skim milk fermented with *Lactobacillus rhamnosus* Bu-Eg6 to 15.25% in skim milk fermented with *Lactobacillus plantarum* Bu-Eg5. These data are confirmed by Santos et al. (2018) and Ismail et al., (2020) reported that the yield of yoghurt after freeze drying was 18% and 16.99%.

3.3. Bulk density, solubility and water holding capacity of fresh freeze-dried skimmed milks and fermented with *Lactobacillus*

3.3.1. The bulk density and solubility index of freeze-dried skimmed milk treatments

The Bulk density of fresh freeze-dried skimmed milks and fermented with *Lactobacillus* was 16.67 g/mL. and there were no changes in bulk density of all treatments of fresh and freeze-dried fermented skim milks with *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6 under investigation. The solubility index of fresh freeze-dried skimmed milk (C) was 100%, followed by 78.95% for skimmed milk fermented with *Lactobacillus plantarum* Bu-Eg5 and fermented with a 1:1 mixture of *Lactobacillus plantarum* Bu-Eg5 (T₂) and *Lactobacillus rhamnosus* Bu-Eg6 (T₃). The lowest solubility was 75% for the freeze-dried skimmed milk fermented with *Lactobacillus rhamnosus* Bu-Eg6 (T₂) and Ismail et al., (2020) found that the average bulk density of freeze-dried yoghurt powders was 1.475 g/mL. and the highest solubility index of freeze-dried yoghurt reached to 86.01%, 84.4 and 84.35% for yoghurts. The changes in in bulk density and solubility index of freeze-dried skimmed milk treatments may be due to differences in structures of treatments. Akash and Yugal (2017) reported that fermented powder had bulk density than milk powder.

3.3.2. The water holding capacity (WHC) of freeze-dried skimmed milk treatments

Water holding capacities (WHC) of freeze-dried skimmed milks are shown in Table 3. WHC in all fresh freeze-dried skimmed milks was 11.16% lower than in skimmed milks fermented with *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6. Differences in WHC are due to differences between fresh freeze-dried skimmed milk and skimmed milks fermented with *Lactobacillus* structures. This reduction in WHC of freeze-dried skimmed milk (control sample) may be due to the high water trapping in the protein matrix of it resulted from its higher total solids content compared to skimmed milks fermented with *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6. These foundations confirmed with those mentioned with Ismail et al., (2020) and Akalin et al., (2012) reported yoghurts have high WHC fortified with whey protein concentrate (WPC).

3.3.3. pH values of freeze-dried skimmed milks fermented with *Lactobacillus*

The pH values of all treatments were decreased after 24 h of fermentation (Table 4), this reflects the increase in acidity and the conversion of lactose to lactic acid by the used microorganisms. The pH of the control recorded 6.57 initially and 6.50 after 24 h of incubation at 37 °C. Mixed ANOVA statistical analysis of pH data showed that, pH levels decreased over the fermentation period in all treatments, but less in the control. There was a statistically significant interaction between the intervention and time on the pH levels $p < 0.0005$ but not for the control. The main effect of treatment on the pH levels showed significant statistical differences.

3.4. Amino acids content of freeze-dried skimmed milk

The effect of different ratio of *Lactobacillus* added on essential and non-essential amino acids content of freeze-dried skimmed milk treatments is shown in Table 4 and Figs. 1–4. In various freeze-dried skimmed milk samples, the major essential amino acid was Leucine acid (3.28%), followed by Lysine acid (3.04%) but Valine acid content (2.25%) was the lowest in freeze-dried fresh skimmed milk sample. On the other hand, the highest non-essential amino acids was Glutamic acid (7.05%), followed by Proline acid (3.16%) and Aspartic acid (2.63%). On the contrary, Cystine acid (0.17%) had the lowest content than the other amino acids. From the same Table 4 the highest total essential amino acids were observed in freeze-

Table 3 Bulk density, solubility, water holding capacity and pH values of freeze-dried skimmed milks fermented with *Lactobacillus*.

Treatments	Bulk density (g/mL.)	Solubility (%)	Water holding capacity (WHC %)	pH values	
				Zero time	After 24 h
C	16.67	100	11.16	6.57 ³	6.50 ⁴
T ₁	16.67	78.95	30.23	6.48 ¹	4.11 ¹
T ₂	16.67	75	28.37	6.51 ^{1,2}	4.42 ³
T ₃	16.67	78.95	31.16	6.54 ^{2,3}	4.32 ²

C: Control (Fresh freeze-dried skimmed milk); T₁: skimmed milk fermented with *Lactobacillus plantarum* Bu-Eg5; T₂: freeze-dried skimmed milk fermented with *Lactobacillus rhamnosus* Bu-Eg6; T₃: freeze-dried skimmed milk fermented with 1:1 *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6.

Table 4 Amino acids content of freeze-dried skimmed milks fermented with *Lactobacillus*.

Amino acids (%)		Control	T ₁	T ₂	T ₃
Essential amino acids					
Therionine	(Thr)	1.47	1.50	1.40	1.03
Valine	(Val)	2.25	2.09	2.11	1.72
Methionine	(Met)	0.00	0.00	0.00	0.00
Isoleucine	(Ile)	1.84	1.95	1.93	1.46
Leucine	(Leu)	3.28	3.15	3.16	2.53
Tyrosine	(Tyr)	2.23	1.96	1.97	1.49
Phenalanine	(Phe)	2.10	1.85	1.86	1.42
Lysine	(Lys)	3.04	2.73	2.72	2.36
Histidine	(His)	1.05	0.99	1.00	0.86
Total essential amino acids		17.26	16.22	16.15	12.87
Non-essential amino acids					
Aspartic	(Asp)	2.63	2.61	2.55	2.13
Serine	(Ser)	1.81	1.89	1.67	1.24
Glutamic	(Glu)	7.05	6.75	6.69	5.42
Proline	(Pro)	3.16	3.23	3.25	3.18
Glycine	(Gly)	0.66	0.63	0.63	0.54
Alanine	(Ala)	0.98	0.99	0.99	0.75
Cystine	(Cys)	0.17	0.04	0.31	0.61
Arginine	(Arg)	1.18	1.10	1.14	0.86
Total non-essential amino acids		17.64	17.24	17.23	14.73
Total amino acid		34.9	33.46	33.38	27.60

C: Control (Fresh freeze-dried skimmed milk); T₁: skimmed milk fermented with *Lactobacillus plantarum* Bu-Eg5; T₂: freeze-dried skimmed milk fermented with *Lactobacillus rhamnosus* Bu-Eg6; T₃: freeze-dried skimmed milk fermented with 1:1 *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6.

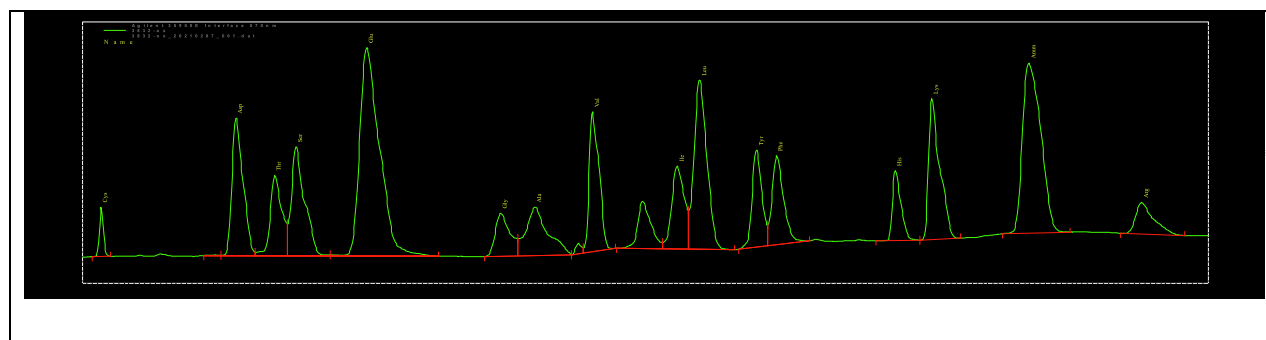
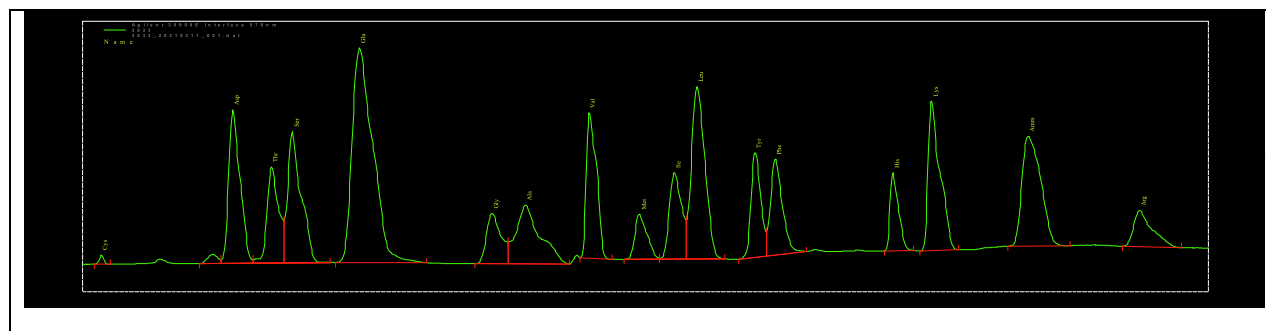
**Fig. 3** Chromatogram of amino acids content of freeze-dried fermented milks with *Lactobacillus rhamnosus* Bu-Eg6.**Fig. 4** Chromatogram of amino acids content of freeze-dried fermented milks with 1:1 *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6.

Table 5 Microbiological evaluation of freeze-dried skimmed milks fermented with *Lactobacillus*.

Microbial count type	Log CFU/ ml			
	C	T ₁	T ₂	T ₃
Lactic acid bacteria counts	1.09	7.94	7.68	7.94
Yeast and mould counts	ND*	ND*	ND*	ND*
Coliform counts	ND*	ND*	ND*	ND*

ND* = not detected; C: Control (Fresh skimmed milk); T₁: skimmed milk fermented with *Lactobacillus plantarum* Bu-Eg5; T₂: skimmed milk fermented with *Lactobacillus rhamnosus* Bu-Eg6; T₃: skimmed milk fermented with 1:1 *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6.

dried of fresh skimmed milk sample (17.26%) followed by freeze-dried of skimmed milk sample fermented with *Lactobacillus plantarum* Bu-Eg5 (16.22%) while the lowest recorded in freeze-dried skimmed milk sample fermented with 1:1 *Lactobacillus plantarum* Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6 (12.87%). It could also be noticed that *Lactobacillus* need sufficient amount from essential and non-essential amino acids to survive, moreover inoculation of *Lactobacillus plantarum* Bu-Eg5 with *Lactobacillus rhamnosus* Bu-Eg6 to ferment skim milk take or consumption more amounts of amino acids compared with *Lactobacillus plantarum* Bu-Eg5 or *Lactobacillus rhamnosus* Bu-Eg6 only, these results may be due to the competitiveness to life between them.

3.5. Microbiological evaluation of skimmed milks fermented

Lactic acid bacteria counts, yeast and mould counts and coliform counts of skimmed milks fermented tabulated in table 5. Despite the heat treatment of skim milk (121 °C for 5 min), there was a little count in the control treatment (1.09 log CFU/ml). Yeast & mould and coliforms were not detected due to the sanitary conditions during the work.

4. Conclusion

Lactobacillus plantarum Bu-Eg5 and *Lactobacillus rhamnosus* Bu-Eg6 were used to ferment skim cow milk and a laboratory freeze dryer used as freeze-drying fresh and fermented milks. Powders were evaluated for their solubility index, bulk density, water holding capacity (WHC), chemical composition, amino acids and microbiological counts (Lactic acid bacteria, Yeast and mould counts and coliform counts). The obtained data mentioned that the major essential amino acid was Leucine acid (3.28%) while Glutamic acid (7.05%) was the highest non-essential amino acids followed by Proline acid (3.16%). Despite the heat treatment of skim milk (121 °C for 5 min), there was a little count in the control treatment (1.09 log CFU/ml). Yeast & mould and coliforms were not detected due to the sanitary conditions during the work. The results showed that the total protein content was 38.69% for fresh freeze-dried skimmed milk and this value was higher than the fermented freeze-dried skimmed milks. Bulk density of freeze-dried skimmed milks was 16.67 g/mL. The solubility was 100% for fresh freeze-dried skimmed milks the highest followed by 78.95% for each skimmed milk fermented with *Lactobacillus plantarum* Bu-Eg5 and fer-

mented with 1:1 *Lactobacillus plantarum* Bu-Eg5 (T₂) and *Lactobacillus rhamnosus* Bu-Eg6 (T₃). The water holding capacity of fresh freeze-dried skimmed milks was lower than other samples.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

The authors greatly appreciate the Scientific Research Support Programs of Benha University, Ain Shams University and Mansoura University, Egypt. Also, Faculty of Applied Sciences, Umm Al-Qura University, College of Science and Humanities, Shaqra University, Afif. The authors gratefully acknowledges financial support from Taif University Researchers Supporting Project number (TURSP-2020/135), Taif University, Taif, Saudi Arabia.

References

- Akaln, A.S., Unal, G., Dinkci, N., Hayaloglu, A.A., 2012. Microstructural, textural, and sensory characteristics of probiotic yogurts fortified with sodium calcium caseinate or whey protein concentrate. *J. Dairy Sci.* 95 (7), 3617–3628. [https://www.journalofdairyscience.org/article/S0022-0302\(12\)00342-6/fulltext](https://www.journalofdairyscience.org/article/S0022-0302(12)00342-6/fulltext).
- Akash, K., Yugal, A., 2017. Study of quality of milk powder in sterling agro industries limited- nova. *Acta Chemia Malaysia* 1 (2), 8–10. <https://www.actachemicamalaysia.com/archives/2acmy2017/2acmy2017-08-10.pdf>.
- Aly, A.A., El-Deeb, F.E., Abdelazeem, A.A., Hameed, A.M., Abdulaziz Alfi, A., Alessa, H., Alrefaei, A.F., 2021. Addition of whole barley flour as a partial substitute of wheat flour to enhance the nutritional value of biscuits. *Arab. J. Chem.* 14 (5), 103112. <https://doi.org/10.1016/j.arabjc.2021.103112>.
- Aly, A.A., Refaey, M.M., Hameed, A.M., Sayqal, A., Abdella, S.A., Mohamed, A.S., Hassan, M.A., Ismail, H.A., 2020. Effect of addition sesame seeds powder with different ratio on microstructural and some properties of low fat Labneh. *Arab. J. Chem.* 13 (10), 7572–7582. <https://doi.org/10.1016/j.arabjc.2020.08.032>.
- AOAC, 2019. Official Methods of Analysis (OMA). Association of Official Analytical Chemists. Official Methods of Analysis. AOAC, Washington, DC, USA. <https://www.aoac.org/official-methods-of-analysis-21st-edition-2019/>.
- Carvalho, A.S., Silva, J., Ho, P., Teixeira, P., Malcata, F.X., Gibbs, P., 2004. Relevant factors for the preparation of freeze-dried lactic acid

- bacteria. *Int. Dairy J.* 14 (10), 835–847. <https://doi.org/10.1016/j.idairyj.2004.02.001>.
- Chen, H., Chen, S., Li, C., Shu, G., 2015. Response surface optimization of lyoprotectant for *Lactobacillus bulgaricus* during vacuum freeze-drying. *Preparat. Biochem. Biotechnol.* 45 (5), 463–475. <https://doi.org/10.1080/10826068.2014.923451>.
- Conrad, P.B., Miller, D.P., Cielenski, P.R., de Pablo, J.J., 2000. Stabilization and preservation of *Lactobacillus acidophilus* in saccharide matrices. *Cryobiology* 41 (1), 17–24. <https://pubmed.ncbi.nlm.nih.gov/11017757/>.
- Domínguez Díaz, L., Fernández-Ruiz, V., Cámara, M., 2020. An international regulatory review of food health-related claims in functional food products labeling. *J. Funct. Foods* 68, 103896. <https://doi.org/10.1016/j.jff.2020.103896>.
- FAO/WHO Working Group, 2002. Guidelines for the evaluation of probiotics in food. FAO/WHO Working Group, pp. 1–11.
- Gul, L.B., Con, A.H., Gul, O., 2020. Storage stability and sourdough acidification kinetic of freeze-dried *Lactobacillus curvatus* N19 under optimized cryoprotectant formulation. *Cryobiology* 96, 122–129. <https://pubmed.ncbi.nlm.nih.gov/32712072/>.
- Han, L., Pu, T., Wang, X., Liu, B., Wang, Y., Feng, J., Zhang, X., 2018. Optimization of a protective medium for enhancing the viability of freeze-dried *Bacillus amyloliquefaciens* B1408 based on response surface methodology. *Cryobiology* 81, 101–106. <https://pubmed.ncbi.nlm.nih.gov/29458043/>.
- Hosseini, S.V., Arlindo, S., Böhme, K., Fernández-No, C., Calo-Mata, P., Barros-Velázquez, J., 2009. Molecular and probiotic characterization of bacteriocin-producing *Enterococcus faecium* strains isolated from nonfermented animal foods. *J Appl Microbiol* 107 (4), 1392–1403. <https://pubmed.ncbi.nlm.nih.gov/19426265/>.
- Hubalek, Z., 2003. Protectants used in the cryopreservation of microorganisms. *Cryobiology* 46 (3), 205–229. <https://pubmed.ncbi.nlm.nih.gov/12818211/>.
- Ismail, E.A., Aly, A.A., Atallah, A.A., 2020a. Quality and microstructure of freeze-dried yoghurt fortified with additives as protective agents. *Heliyon* 6 (10). <https://doi.org/10.1016/j.heliyon.2020.e05196>.
- Ismail, H.A., Hameed, A.M., Refaey, M.M., Sayqal, A., Aly, A.A., 2020b. Rheological, physio-chemical and organoleptic characteristics of ice cream enriched with Doum syrup and pomegranate peel. *Arab. J. Chem.* (10), 7346–7356. <https://doi.org/10.1016/j.arabjc.2020.08.012>.
- Lu, Y., Huang, L., Yang, T., Lv, F., Lu, Z., 2017. Optimization of a cryoprotective medium to increase the viability of freeze-dried *Streptococcus thermophilus* by response surface methodology. *LWT* 80, 92–97. <https://www.ars.usda.gov/research/publications/publication/?seqNo115=333535>.
- Martos, G., Minahk, C., Font de Valdez, G., Morero, R., 2007. Effects of protective agents on membrane fluidity of freeze-dried *Lactobacillus delbrueckii* ssp. *bulgaricus*. *Lett. Appl. Microbiol.* 45 (3), 282–288. <https://doi.org/10.1111/j.1472-765X.2007.02188.x>.
- Meisel, H., Bockelmann, W., 1999. Bioactive peptides encrypted in milk proteins: proteolytic activation and thropho-functional properties. In: Konings, W.N., Kuipers, O.P., In't Veld, J.H.J.H. (Eds.), *Lactic Acid Bacteria: Genetics, Metabolism and Applications: Proceedings of the Sixth Symposium on lactic acid bacteria: genetics, metabolism and applications*, 19–23 September 1999, Veldhoven, The Netherlands. Dordrecht: Springer Netherlands, pp. 207–215. <https://pubmed.ncbi.nlm.nih.gov/10532380/>.
- Niu, X., Deng, L., Zhou, Y., Wang, W., Yao, S., Zeng, K., 2016. Optimization of a protective medium for freeze-dried *Pichia membranifaciens* and application of this biocontrol agent on citrus fruit. *J. Appl. Microbiol.* 121 (1), 234–243. <https://doi.org/10.1111/jam.13129>.
- Oberman, H., Libudzisz, Z., 1998. Fermented milks. In: Wood, B.J.B. (Ed.), *Microbiology of Fermented Foods*. Springer, US, Boston, MA, pp. 308–350.
- Pederson, C.S., 1971. *Microbiology of Food Fermentations*. AVI Publishing Co., Inc., Westport, Connecticut, USA.
- Santos, G.d., Pagani, A.A.C., Rosenthal, A., Nunes, T.P., Silva, M.A. A.P.d., 2018. Development and acceptance of freeze-dried yogurt “powder yogurt”. https://www.researchgate.net/publication/326836982_Development_and_acceptance_of_freeze-dried_yogurt_powder_yogurt.
- Stefanello, R.F., Nabeshima, E.H., Iamanaka, B.T., Ludwig, A., Fries, L.L.M., Bernardi, A.O., Copetti, M.V., 2019. Survival and stability of *Lactobacillus fermentum* and *Wickerhamomyces anomalus* strains upon lyophilisation with different cryoprotectant agents. *Food Res. Int.* 115, 90–94. <https://pubmed.ncbi.nlm.nih.gov/30599986/>.
- Suez, J., Zmora, N., Segal, E., Elinav, E., 2019. The pros, cons, and many unknowns of probiotics. *Nat. Med.* 25 (5), 716–729. <https://pubmed.ncbi.nlm.nih.gov/31061539/>.
- Velly, H., Fonseca, F., Passot, S., Delacroix-Buchet, A., Bouix, M., 2014. Cell growth and resistance of *Lactococcus lactis* subsp. *lactis* TOMSC161 following freezing, drying and freeze-dried storage are differentially affected by fermentation conditions. *J. Appl. Microbiol.* 117 (3), 729–740. <https://pubmed.ncbi.nlm.nih.gov/24935668/>.